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## Revisions to the staging and classification of mycosis fungoides and Sézary syndrome: a proposal of the International Society for Cutaneous Lymphomas (ISCL) and the cutaneous lymphoma task force of the European Organization of Research and Treatment of Cancer (EORTC)

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**The ISCL/EORTC recommends revisions to the Mycosis Fungoides Cooperative Group classification and staging system for cutaneous T-cell lymphoma (CTCL). These revisions are made to incorporate advances related to tumor cell biology and diagnostic techniques as pertains to mycosis fungoides (MF) and Sézary syndrome (SS) since the 1979 publication of the original guidelines, to clarify certain**

**variables that currently impede effective interinstitution and interinvestigator communication and/or the development of standardized clinical trials in MF and SS, and to provide a platform for tracking other variables of potential prognostic significance. Moreover, given the difference in prognosis and clinical characteristics of the non-MF/non-SS subtypes of cutaneous lymphoma, this revision per-**

**tains specifically to MF and SS. The evidence supporting the revisions is discussed as well as recommendations for evaluation and staging procedures based on these revisions. (Blood. 2007;110:1713-1722)**

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### Purpose of revision

When it became recognized that mycosis fungoides (MF), Sézary syndrome (SS), and other cutaneous T-cell lymphomas arising in the skin were part of a broader spectrum of cutaneous T-cell lymphoma (CTCL),<sup>1</sup> the Mycosis Fungoides Cooperative Group (MFCG) developed a staging system for CTCL<sup>2</sup> aimed at the specific findings in the MF/SS subtypes and based on the TNM (tumor-node-metastasis) classification advocated by the International Union Against Cancer (UICC)<sup>3</sup> and American Joint Committee on Cancer.<sup>4</sup> This classification and staging system was modified in conjunction with the National Cancer Institute (NCI) and the Veteran's Administration (VA) Hospital and published in 1979 (Tables 1,2).<sup>5</sup> The MFCG system has proved to be an extremely useful tool in the management of patients with MF/SS and is the standard for the staging and classification of MF/SS patients today.

Since the publication of the MFCG classification and staging system, there have been steady advances in the areas of molecular biology, immunohistochemistry, and imaging as well as new data

on prognostic variables in MF and SS that affect staging. In addition, it has become clear that the non-MF/non-SS subtypes of CTCL neither share the same T or N stages nor have the same prognosis as MF and SS.<sup>6,7</sup> The purpose of revising the MFCG staging and classification system now is to incorporate these advances and new data; to exclude the non-MF/non-SS variants; to provide for identification, subsequent tracking, and validation of certain variables that appear to have prognostic importance; to provide clear definitions of certain variables necessary to carry out the staging and classification that in the current system have been open to variations in interpretation; and to incorporate blood (B) involvement, a major prognostic factor for patients with MF and SS.

To address these issues, the International Society for Cutaneous Lymphomas (ISCL), which includes key leaders of the EORTC, sponsored a series of workshops in 2002 to 2006 on the classification and staging of MF and SS, and the resulting revision to the

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**Table 1. Original Mycosis Fungoides Cooperative Group TNM classification of cutaneous T-cell lymphoma (CTCL)**

Classification	Description
<b>T: Skin*</b>	
T <sub>0</sub>	Clinically and/or histopathologically suspicious lesions
T <sub>1</sub>	Limited plaques, papules, or eczematous patches covering < 10% of the skin surface
T <sub>2</sub>	Generalized plaques, papules, or erythematous patches covering ≥ 10% or more of the skin surface
T <sub>3</sub>	Tumors, one or more
T <sub>4</sub>	Generalized erythroderma
<b>N: Lymph nodes†</b>	
N <sub>0</sub>	No clinically abnormal peripheral lymph nodes; pathology negative for CTCL
N <sub>1</sub>	Clinically abnormal peripheral lymph nodes; pathology negative for CTCL
N <sub>2</sub>	No clinically abnormal peripheral lymph nodes; pathology positive for CTCL
N <sub>3</sub>	Clinically abnormal peripheral lymph nodes; pathology positive for CTCL
<b>B: Peripheral blood</b>	
B <sub>0</sub>	Atypical circulating cells not present (< 5%)
B <sub>1</sub>	Atypical circulating cells present (> 5%); record total white blood count and total lymphocyte counts, and number of atypical cells/100 lymphocytes
<b>M: Visceral organs</b>	
M <sub>0</sub>	No visceral organ involvement
M <sub>1</sub>	Visceral involvement (must have pathology confirmation and organ involved should be specified)

\*Pathology of T<sub>1-4</sub> is diagnostic of a CTCL. When more than 1 T exists, both are recorded and the highest is used for staging (eg, T<sub>4(3)</sub>).

†Record number of sites of abnormal nodes (eg, cervical; left + right), axillary (left + right), inguinal (left + right), epitrochlear, and submandibular/submaxillary).

MFCG classification and staging represents a consensus of both groups. The updated ISCL/EORTC staging and classification applies specifically and solely to MF and SS and has maintained the primary components of the MFCG system to allow for continued comparison of patient outcomes within both systems.

### Establishment of the diagnosis of MF/SS

MF and SS represent approximately 65% of the cases of CTCL.<sup>6</sup> Both are characterized by a monoclonal proliferation of predominantly CD4<sup>+</sup>/CD45RO<sup>+</sup> helper T cells and the loss of mature T-cell antigens in the skin and other involved organs.<sup>6,8,9</sup> SS is currently defined by the ISCL as a distinctive erythrodermic CTCL (albeit potentially lacking the diagnostic histologic features in the skin<sup>10</sup>) with hematologic evidence of leukemic involvement.<sup>11</sup> The WHO/

**Table 2. Original Mycosis Fungoides Cooperative Group staging system for cutaneous T-cell lymphoma (CTCL)**

Clinical stage	T	N	M
IA	1	0	0
IB	2	0	0
IIA	1,2	1	0
IIB	3	0,1	0
III	4	0,1	0
IVA	1-4	2,3	0
IVB	1-4	0-3	1

The "B" classification does not alter clinical stage.

EORTC considers SS to be a separate entity from cases that otherwise meet the criteria for SS but have been preceded by clinically typical MF.<sup>6,9</sup> Such latter cases have been designated as "SS preceded by MF" and also as "secondary" SS.<sup>12</sup>

In some instances, the diagnosis of MF can be rendered with confidence on a skin biopsy specimen based on typical light microscopic changes, that is, marked epidermotropism of cytologically atypical T lymphocytes, clusters of these cells in the epidermis (Pautrier microabscesses), or a bandlike infiltrate containing abnormal lymphocytes in the upper dermis.<sup>13-16</sup> However, a definitive histopathologic diagnosis by light microscopy alone may be difficult to make in early MF<sup>17,18</sup> or in erythroderma in which inflammatory cells often predominate.<sup>19</sup> The ISCL recently proposed a diagnostic algorithm for early MF (Table 3).<sup>15</sup> No patient with clinically suspect patch- or plaque-stage disease who does not at least fulfill this algorithm nor any patient with tumor-stage disease with histologic findings only "suggestive of MF" should be entered into MF/SS databases or into therapeutic trials for MF/SS. In the case of erythrodermic CTCL, multiple skin biopsies may be necessary to establish a firm diagnosis or a definitive diagnosis may be made by blood studies and/or by biopsy of an enlarged lymph node or other tissue.<sup>19,20</sup>

For patients presenting with tumors, it is important to differentiate tumor-stage MF from non-MF subtypes of CTCL. In classic MF, the tumor lesions generally develop in the presence of patch or plaque disease and not de novo. In the past, this latter presentation was classified as MF tumor d'emblee, but many of these cases would now, with immunophenotypic markers, likely be classified as various types of non-MF T-cell lymphoma or even B-cell lymphoma of the skin.<sup>6,7</sup> Given the large number of neoplastic lymphocytes in tumoral lesions of CTCL, molecular analysis of tumor lesions will usually, but not invariably,<sup>21</sup> demonstrate evidence of a T-cell clone; however, if this is lacking, chromosomal analysis<sup>22</sup> and/or additional molecular studies to rule out a T-cell-rich B-cell lymphoma are in order. Histologic evaluation of an enlarged node can also be used to confirm the type of non-MF CTCL and/or to confirm (and stage) MF.

**Table 3. Algorithm for the diagnosis of early MF<sup>15</sup>**

Criteria	Major (2 points)	Minor (1 point)
<b>Clinical</b>		
Persistent and/or progressive patches and plaques plus	Any 2	Any 1
(1) Non-sun-exposed location		
(2) Size/shape variation		
(3) Poikiloderma		
<b>Histopathologic</b>		
Superficial lymphoid infiltrate plus	Both	Either
(1) Epidermotropism without spongiosis		
(2) Lymphoid atypia*		
Molecular/biologic: clonal TCR gene rearrangement	NA†	Present
<b>Immunopathologic</b>		
(1) CD2,3,5 less than 50% of T cells	NA†	Any 1
(2) CD7 less than 10% of T cells		
(3) Epidermal discordance from expression of CD2,3,5 or CD7 on dermal T cells		

— indicates not applicable.

\*Lymphoid atypia is defined as cells with enlarged hyperchromatic nuclei and irregular or cerebriform nuclear contours.

†Not applicable since it cannot fulfill any major criteria.

**Table 4. ISCL/EORTC revision to the classification of mycosis fungoides and Sézary syndrome**

TNMB stages	
<b>Skin</b>	
T <sub>1</sub>	Limited patches,* papules, and/or plaques† covering < 10% of the skin surface. May further stratify into T <sub>1a</sub> (patch only) vs T <sub>1b</sub> (plaque ± patch).
T <sub>2</sub>	Patches, papules or plaques covering ≥ 10% of the skin surface. May further stratify into T <sub>2a</sub> (patch only) vs T <sub>2b</sub> (plaque ± patch).
T <sub>3</sub>	One or more tumors‡ (≥ 1-cm diameter)
T <sub>4</sub>	Confluence of erythema covering ≥ 80% body surface area
<b>Node</b>	
N <sub>0</sub>	No clinically abnormal peripheral lymph nodes§; biopsy not required
N <sub>1</sub>	Clinically abnormal peripheral lymph nodes; histopathology Dutch grade 1 or NCI LN <sub>0,2</sub>
N <sub>1a</sub>	Clone negative#
N <sub>1b</sub>	Clone positive#
N <sub>2</sub>	Clinically abnormal peripheral lymph nodes; histopathology Dutch grade 2 or NCI LN <sub>3</sub>
N <sub>2a</sub>	Clone negative#
N <sub>2b</sub>	Clone positive#
N <sub>3</sub>	Clinically abnormal peripheral lymph nodes; histopathology Dutch grades 3-4 or NCI LN <sub>4</sub> ; clone positive or negative
N <sub>x</sub>	Clinically abnormal peripheral lymph nodes; no histologic confirmation
<b>Visceral</b>	
M <sub>0</sub>	No visceral organ involvement
M <sub>1</sub>	Visceral involvement (must have pathology confirmation¶) and organ involved should be specified)
<b>Blood</b>	
B <sub>0</sub>	Absence of significant blood involvement: ≤ 5% of peripheral blood lymphocytes are atypical (Sézary) cells
B <sub>0a</sub>	Clone negative#
B <sub>0b</sub>	Clone positive#
B <sub>1</sub>	Low blood tumor burden: > 5% of peripheral blood lymphocytes are atypical (Sézary) cells but does not meet the criteria of B <sub>2</sub>
B <sub>1a</sub>	Clone negative#
B <sub>1b</sub>	Clone positive#
B <sub>2</sub>	High blood tumor burden: ≥ 1000/μL Sézary cells   with positive clone#

\*For skin, patch indicates any size skin lesion without significant elevation or induration. Presence/absence of hypo- or hyperpigmentation, scale, crusting, and/or poikiloderma should be noted.

†For skin, plaque indicates any size skin lesion that is elevated or indurated. Presence or absence of scale, crusting, and/or poikiloderma should be noted. Histologic features such as folliculotropism or large-cell transformation (> 25% large cells), CD30<sup>+</sup> or CD30<sup>-</sup>, and clinical features such as ulceration are important to document.

‡For skin, tumor indicates at least one 1-cm diameter solid or nodular lesion with evidence of depth and/or vertical growth. Note total number of lesions, total volume of lesions, largest size lesion, and region of body involved. Also note if histologic evidence of large-cell transformation has occurred. Phenotyping for CD30 is encouraged.

§For node, abnormal peripheral lymph node(s) indicates any palpable peripheral node that on physical examination is firm, irregular, clustered, fixed or 1.5 cm or larger in diameter. Node groups examined on physical examination include cervical, supraclavicular, epitrochlear, axillary, and inguinal. Central nodes, which are not generally amenable to pathologic assessment, are not currently considered in the nodal classification unless used to establish N<sub>3</sub> histopathologically.

¶For viscera, spleen and liver may be diagnosed by imaging criteria.

||For blood, Sézary cells are defined as lymphocytes with hyperconvoluted cerebriform nuclei. If Sézary cells are not able to be used to determine tumor burden for B<sub>2</sub>, then one of the following modified ISCL criteria along with a positive clonal rearrangement of the TCR may be used instead: (1) expanded CD4<sup>+</sup> or CD3<sup>+</sup> cells with CD4/CD8 ratio of 10 or more, (2) expanded CD4<sup>+</sup> cells with abnormal immunophenotype including loss of CD7 or CD26.

#A T-cell clone is defined by PCR or Southern blot analysis of the T-cell receptor gene.

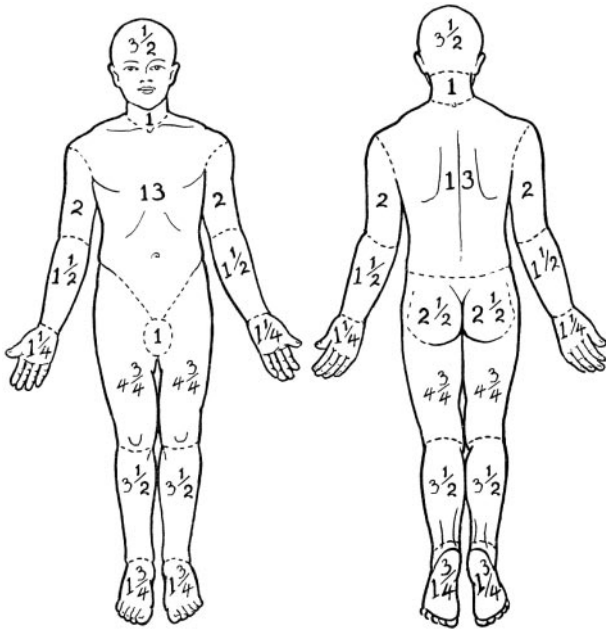
## Proposed revisions to the T (skin) classification

The original MFCG skin scoring system for CTCL (Table 1) included a T<sub>0</sub> category for “clinically and/or histopathologically suspicious lesions.” While T<sub>0</sub> may be a useful category for tracking disorders with malignant potential, current practice dictates that clinical staging be applied only to cases in which a diagnosis of cancer has been established. Therefore, the T<sub>0</sub> category has been eliminated in the ISCL/EORTC updated staging and classification scheme (Table 4) that requires all staged patients have a definitive diagnosis of MF/SS and/or algorithmic diagnosis of early MF.

The definition and differentiation of patch versus plaque<sup>23</sup> versus tumor lesions<sup>24,25</sup> is more than of just semantic importance because both prognosis and choice of and response to treatment are linked to these different designations. Currently, the distinction between patch and thin plaque lesions and between thick plaque and tumor lesions in MF is quite subjective<sup>4,26</sup> (personal communication, E. Vonderheid, ISCL Workshop San Francisco, 1999). The ISCL/EORTC have attempted to add some clarity to the situation by suggesting definitions for the skin lesions (Table 4). Because in patch/plaque disease, histology has been shown to offer an

objective means of defining each subtype,<sup>27,28</sup> be a validated surrogate for the clinical classification of MF lesions,<sup>29</sup> and have prognostic implications,<sup>30</sup> there is a provision in the classification system for characterizing exclusively patch-stage disease with the subscript of “a” (T<sub>1a</sub> and T<sub>2a</sub>) versus combined patch/plaque disease with the subscript of “b” (T<sub>1b</sub> and T<sub>2b</sub>) in order to gather additional longitudinal data on this distinction (Table 4). However, the derivation of all T stages remains a clinical determination.

In both the original MFCG<sup>5</sup> and the revised staging system, the T<sub>1</sub> skin rating is defined as papules, patches, and/or plaques covering less than 10% body surface area (BSA) and T<sub>2</sub> skin rating is defined as patches and/or plaques covering 10% or more BSA. In the classification published by the MFCG in 1979, 1% BSA was defined as equal to the “palmar surface of the hand.”<sup>5</sup> However, the area of the palm and digits together is actually slightly less than 1% BSA (~ 0.8%),<sup>31-34</sup> and mathematically and reliably, the palm at 0.5% BSA<sup>31,34</sup> may be the easiest and most reliable measure to use in assigning BSA of lesions of MF or SS. Another method of determining BSA is to estimate the percentage of skin involvement in each of 12 regions of the body (each with a relative assigned percent BSA<sup>35</sup> [Figure 1]), multiplying this number by the percentage of the BSA for that particular region and adding up the regional percentages to obtain the total BSA involved with MF/SS.



**Figure 1.** Regional percent body surface area (BSA) in the adult. Adapted from Lund and Browder<sup>35</sup> with permission.

Although ulceration, which may occur in plaques as well as tumors, is generally a poor prognostic sign,<sup>4,36,37</sup> ulceration may be caused by infection as well as tumor necrosis and, by multivariable analysis, does not alter the prognosis once the extent of the T rating is known.<sup>38</sup> Therefore, the ISCL/EORTC does not recommend using ulceration as the sole criterion to move a patient from plaque (T<sub>1</sub> or T<sub>2</sub>)- to tumor (T<sub>3</sub>)-stage MF.

Whether any given number of lesions, aggregate volume, size of largest lesion, number of lesions, or specific body regions involved has any predictive prognostic value in tumor-stage MF is unknown at this time. The MFCG originally required that a diagnosis of tumor-stage disease include at least 3 tumors,<sup>26</sup> but this was changed to one or more tumors in the final MFCG staging system.<sup>5</sup> The proposed ISCL/EORTC classification revision retains the requirement of at least one tumor ( $\geq 1.5$  cm in diameter) for the definition of T<sub>3</sub>. Whether there should be a minimum histologic depth of infiltrate to distinguish plaque from tumor in order to corroborate this important assignment of T stage based on a single lesion has not been yet been determined.

The skin of erythrodermic CTCL may show some degree of clinically apparent infiltration that may be caused by either dermal infiltration with tumor cells or an inflammatory reaction with or without edema. There is currently no distinction in the updated staging system for subclassifying T<sub>4</sub> based on varying degrees of induration, erythema, or scale. Specific grading systems for erythroderma that do include these variables have been published elsewhere<sup>39,40</sup> and may be of value for use in clinical trials.

When more than one T rating might apply, the highest is used for staging purposes. In the situations where both tumors and erythroderma exist simultaneously, both T ratings should be recorded (eg, T<sub>4(3)</sub>).<sup>5</sup> This latter nuance of the T staging suggested by the MFCG continues to offer a way of tracking a variable that may impact on the poor prognosis of T<sub>4</sub> disease, but would otherwise be buried in the classification hierarchy.

There are at least 2 histologic findings on skin biopsies in MF that appear to have prognostic importance but require further data before

modifications to the staging system are justified. Folliculotropic MF is characterized histologically by atypical CD4<sup>+</sup> T lymphocytes that surround and infiltrate the hair follicles (folliculotropism), usually without evidence of epidermotropism and with frequent concomitant follicular mucinosis.<sup>41-43</sup> Clinically, folliculotropic MF is typically classified under the T<sub>1</sub> or T<sub>2</sub> skin rating even though the infiltrate extends histologically along the hair follicles deeper than is typical for plaque-stage disease.<sup>41,44</sup> Folliculotropic MF has been shown to be associated with a worse prognosis than expected for clinical stage<sup>41-43,45</sup>: the 5-year survival is similar and the 10-year survival is worse than in patients with tumor-stage MF.<sup>45</sup> Large-cell transformation, defined as a biopsy specimen showing large cells ( $\geq 4$  times the size of a small lymphocyte) in 25% or more of the dermal infiltrate,<sup>46-49</sup> is a poor prognostic sign, seen most commonly in tumor-stage MF and less commonly in plaque-stage and erythrodermic MF. Based on molecular analysis, large-cell transformation in MF or SS represents evolution of the original malignant clone.<sup>50,51</sup> These large cells may or may not be CD30<sup>+</sup>.<sup>48</sup> The possibility that a patient with CD30<sup>+</sup> nodules might have primary cutaneous CD30<sup>+</sup> anaplastic large-cell lymphoma coexisting with MF must also be considered, although the coexistence of typical patches and plaques of MF would normally indicate that such lesions represent large-cell transformation (CD30<sup>+</sup>) of MF rather than a separate primary cutaneous lymphoma. The ISCL/EORTC recommends tracking folliculotropic MF and large-cell transformation to determine if either warrants an advance in stage.

## Revisions to the N (node) classification

### Defining peripheral adenopathy

The negative impact on survival of “palpable adenopathy” in MF has long been appreciated.<sup>14,24,26,36,37,52,53</sup> In the MFCG classification, “N” represented only peripheral lymph nodes. Although there was no size designation for “abnormal” nodes, this was not problematic as all patients, even those without palpable nodes, were to have node biopsies as part of the staging evaluation since each N rating for staging was based on both clinical and histopathologic findings. Although Sausville et al reported that 9 (32%) of 28 patients with MF or SS without adenopathy had either frank lymphoma or advanced dermatopathic findings on histopathologic review of “blind” nodal biopsies,<sup>13</sup> biopsies of nonpalpable nodal groups are rarely done in clinical practice today.

The updated ISCL/EORTC classification eliminates biopsies of lymph nodes for staging purposes that are not enlarged on physical examination or imaging studies. In doing so, however, the size of a peripheral lymph node designated to be “clinically abnormal” takes on more importance because a lymph node biopsy is recommended only of such nodes. The ISCL/EORTC revision defines clinically abnormal peripheral nodes as those 1.5 cm or larger in the longest transverse diameter or any palpable peripheral node, regardless of size, that on physical examination is firm, irregular, clustered, or fixed. The 1.5-cm size is different from the 1-cm diameter node designated as abnormal both by the International Workshop on Response Criteria for Non-Hodgkin Lymphoma<sup>54</sup> and the ISCL/EORTC staging for non-MF/SS primary cutaneous lymphomas<sup>55</sup> since reactive or dermatopathic peripheral lymph nodes commonly occur in MF, SS, and benign inflammatory skin disorders<sup>56</sup> but are uncommon in these other lymphomas involving the skin. These clinically enlarged or abnormal nodes should be corroborated by an imaging study (computed tomography [CT]  $\pm$  <sup>18</sup>F-fluorodeoxyglucose positron emission tomography [FDG-PET] or by magnetic

resonance imaging (MRI) (in cases where patients may be allergic to contrast dye) or ultrasound prior to biopsy.

### Pathology of lymph nodes

The original N rating did not include guidelines for the distinction between nodes judged pathologically “negative” or “positive” for CTCL nor did it give any weight to increasing magnitude of tumor involvement based on light microscopy, flow cytometry, or molecular genetic results. The 2 main histopathologic grading systems for lymph nodes in MF/SS in use today are the NCI/VA classification system,<sup>13</sup> first proposed by Matthews and Gazdar,<sup>57</sup> and the Dutch System<sup>58</sup> (Table 5). The major difference between these classification systems resides in the criteria used to define “abnormal” lymphocytes.<sup>58,60</sup> Specifically, the NCI/VA system,<sup>13</sup> although it defines abnormal (neoplastic) cells as small (6-10  $\mu\text{m}$ ) or large ( $> 11.5 \mu\text{m}$ ) cells with cerebriform, irregularly folded, hyperconvoluted nuclei (ie, Sézary cells), does not use the size of cells but instead uses the relative numbers of such cells in the paracortex of the lymph node for the LN<sub>0-2</sub> definition (Table 5). Conversely, the Dutch system uses the diameter of the cerebriform cells ( $> 7.5 \mu\text{m}$ ) to define abnormal (neoplastic) cells, and if present, this constitutes early involvement (grade 2).<sup>58</sup>

Prognosis in MF/SS is clearly related to partially or completely effaced nodal architecture as defined by either the NCI-VA (LN<sub>4</sub>) or Dutch (grade 3/4) grading system,<sup>60</sup> and continues to define the N<sub>3</sub> node rating in the updated ISCL/EORTC staging system. However, the prognostic importance of lesser degrees of involvement as defined by light microscopy alone is less clear.<sup>24,59,61,62</sup> Some investigators advocate that the LN<sub>3</sub> rating should be considered equivalent to LN<sub>4</sub> for staging purposes.<sup>13</sup> Although in the context of MF and SS, the survival of patients with an NCI/VA LN<sub>3</sub> node rating is worse than patients with LN<sub>0-2</sub>,<sup>24,62</sup> it must be remembered that findings comparable with LN<sub>3</sub> have been found in nodes from patients with benign disorders<sup>56,60,63</sup> and that the survival of patients with LN<sub>4</sub> is worse than for those with LN<sub>3</sub>.<sup>24,60</sup>

### Prognostic significance of T-cell clonality in lymph nodes

As early as 1988, Sausville et al recognized the potential prognostic significance of a clonal rearrangement of the TCR in nodal staging.<sup>24</sup> In 3 studies that have reported on the use of Southern blot technique, which has a 5% detection threshold to show clonal T cells in lymph nodes,<sup>62,64,65</sup> none of the lymph nodes classified histologically as uninvolved (NCI/VA LN<sub>0-1</sub> or Dutch grade 1) had evidence of a clone. However, 13% of LN<sub>2</sub> nodes, 83% of LN<sub>3</sub> nodes, and 100% of patients with LN<sub>4</sub> nodes had clonal rearrangements in one of the studies.<sup>62</sup> These studies indicate that Southern blot analysis is useful as an adjunct study for lymph nodes that show higher histologic grades of involvement, especially given that

the prognosis of such patients with evidence of a clone was worse than patients without a clone.<sup>62</sup>

However, the Southern blot technique has largely been replaced by the technically easier polymerase chain reaction (PCR)-based methods, with much more sensitive detection thresholds.<sup>66</sup> Although not significant in a multivariate analysis, Fraser-Andrews et al<sup>67</sup> have reported detection of T-cell clones by PCR in 6 of 19 N<sub>0-2</sub> lymph nodes and Assaf et al<sup>68</sup> in 7 of 14 dermatopathic lymph nodes in patients with MF, and in none of the lymph nodes obtained from patients with benign conditions. In a comparison of Southern blot and PCR determination of clonality of TCR rearrangement in both palpable and nonpalpable lymph nodes in patients with MF/SS, Juarez et al concurred that in a univariate analysis, both were predictive of a poor outcome but that only Southern blot analysis was predictive of a poor prognosis in a multivariate analysis that included skin stage, presence or absence of lymphadenopathy, and histologic lymph node score.<sup>69</sup>

For staging purposes, the ISCL recommends that nodal rating still be based on histopathology until new molecular markers have been validated, and that uninvolved nodes exhibiting an NCI/VA grade LN<sub>3</sub> or Dutch grade 2 be classified as an N<sub>2</sub> rating with further division into 2 subgroups: N<sub>2a</sub> (clone negative) and N<sub>2b</sub> (clone positive). It is hoped that by capturing this information longitudinally, it can be determined if there is a similar prognosis of patients with N<sub>2b</sub> and N<sub>3</sub> node ratings.

### Clinical-only staging of nodes

Because a biopsy of a clinically abnormal node is not always done at initial staging, the revised ISCL/EORTC classification has added a new category, the N<sub>x</sub> node rating, to facilitate capture of at least this clinical information.

### Lymph node biopsy

The ISCL/EORTC recommends excisional biopsy as the preferred method to evaluate abnormal lymph nodes in MF/SS. In addition to routine histologic examination, a portion of the node can be processed for immunohistochemistry, flow cytometry, and/or molecular genetic or cytogenetic analysis. Since excisional lymph node biopsies put the patient at risk for sepsis, especially in erythrodermic patients whose skin is often colonized with *Staphylococcus*, alternative methods to obtain nodal tissue (core biopsy or fine needle aspiration [FNA]) have been suggested as potential substitutes particularly if combined with flow cytometry.<sup>70</sup> However, these alternate methods may have inadequate or poorly representative sampling<sup>71</sup> or incomplete concordance with excisional biopsy results,<sup>72,73</sup> and they do not provide the histopathologic assessment of nodal architecture necessary for N staging.

**Table 5. Histopathologic staging of lymph nodes in mycosis fungoides and Sézary syndrome**

Updated ISCL/EORTC classification	Dutch system <sup>58</sup>	NCI-VA classification <sup>13,57,59</sup>
N <sub>1</sub>	Grade 1: dermatopathic lymphadenopathy (DL)	LN <sub>0</sub> : no atypical lymphocytes LN <sub>1</sub> : occasional and isolated atypical lymphocytes (not arranged in clusters) LN <sub>2</sub> : many atypical lymphocytes or in 3-6 cell clusters
N <sub>2</sub>	Grade 2: DL; early involvement by MF (presence of cerebriform nuclei $> 7.5 \mu\text{m}$ )	LN <sub>3</sub> : aggregates of atypical lymphocytes; nodal architecture preserved
N <sub>3</sub>	Grade 3: partial effacement of LN architecture; many atypical cerebriform mononuclear cells (CMCs) Grade 4: complete effacement	LN <sub>4</sub> : partial/complete effacement of nodal architecture by atypical lymphocytes or frankly neoplastic cells

In general, the largest peripheral lymph node draining an area of involved skin and/or one that shows intense uptake on an FDG-PET scan should be selected for biopsy. If there are multiple enlarged nodes, the order of preference for biopsy by location remains cervical, axillary, then inguinal<sup>5</sup> since cervical nodes have a higher chance of showing lymphomatous involvement than other sites.<sup>74</sup> A biopsy of multiple nodes of a single nodal group may show different LN histopathologic ratings, raising the issue of “sampling error” and whether more than one node should be sampled at the time of excision.<sup>75</sup> However, the need to study multiple lymph nodes from one region may be obviated if ancillary studies are performed on the specimen.<sup>76</sup>

Potential involvement of central nodes in MF or SS was not addressed in the original MFCG classification.<sup>5</sup> Central adenopathy may be secondary to a second malignancy (especially a second lymphoma), infection, a reactive process, or MF. The ISCL/EORTC recommends that central enlarged nodes be excluded from the determination of “N” status except in cases where an excisional biopsy of a central node has proven lymphomatous (N<sub>3</sub>) involvement with MF.

## Revision to M classification

Visceral involvement with MF/SS is a well-documented, independently significant prognostic factor in MF/SS.<sup>24,77</sup> But what constitutes “visceral disease” has not been well defined. It is generally asymptomatic and therefore prone to be underdiagnosed, especially in those with more advanced skin involvement. Visceral involvement should be questioned in the absence of node or blood involvement. To be considered as having visceral disease (stage IVb), documentation of involvement by only one organ outside the skin, nodes, or blood is needed.

Splenomegaly in MF patients is generally proven to be either diffuse or nodular involvement with MF<sup>14</sup> and is uncommon in healthy persons or in those with benign skin disease. The ISCL/EORTC considers splenomegaly as visceral disease, even without biopsy confirmation, when it is (a) unequivocally present on physical exam and (b) documented radiographically by either enlargement or multiple focal defects that are neither cystic nor vascular (a more rigorous modification of the Cotswolds meeting recommendations on definition of lymphomatous involvement of the spleen in Hodgkin lymphoma).<sup>78</sup>

Liver disease may be suspected by physical examination, abnormal liver function tests, or radiologic tests (CT, FDG-PET, liver/spleen scan) but should be confirmed by liver biopsy.<sup>79</sup> In agreement with the Cotswolds meeting on Hodgkin lymphoma, multiple focal hepatic defects, which are neither cystic nor vascular, on at least 2 imaging techniques may be considered indicative of tumor involvement.<sup>78</sup>

Bone marrow biopsy confirmation of frank lymphoma is a low-yield procedure in MF unless there is evidence of blood or nodal disease.<sup>24,38,47,52,61,80</sup> Although it has been suggested that the finding of cytologic atypical lymphoid aggregates in the bone marrow of patients with MF correlates with shorter survival,<sup>24,81-83</sup> multivariate analysis has not demonstrated the independent prognostic value of bone marrow involvement.<sup>23,80,84,85</sup> The ISCL/EORTC recommends performance of a bone marrow biopsy in patients with MF and SS who have B<sub>2</sub> blood involvement (as described in paragraph 4 of “Revisions to the B (blood) rating”) or unexplained hematologic abnormalities. Tracking and recording the specific

cytologic findings and whether clonality of the TCR gene rearrangement is present will provide further data on whether bone marrow involvement has independent prognostic significance and should be considered as evidence of visceral disease.

If lung abnormalities or other suggestions of extracutaneous lymphomatous involvement besides splenomegaly are found on radiographic examination, pathological assessment is recommended before ascribing this to visceral involvement with MF/SS. Visceral abnormalities seen radiologically in MF could be secondary to either another unrelated cancer (second malignancies are not uncommon in MF/SS<sup>86-88</sup>) or to an infectious disorder and not to MF/SS.

## Revisions to the B (blood) rating

Following the lead of Clendenning et al,<sup>89</sup> the original MFCG classification of blood involvement in MF (B<sub>1</sub> rating) was defined as more than 5% of the total lymphocytes that exhibited an atypical convoluted appearance by light microscopy.<sup>5</sup> Low numbers of atypical cerebriform mononuclear cells have been reported in both benign skin conditions<sup>90-94</sup> and in healthy donors,<sup>92,95</sup> and can be generated *in vivo* by incubating lymphocytes with cellular mitogens<sup>96</sup> or stimulating peripheral T cells via CD3 or CD2 in the presence of phorbol esters.<sup>97</sup> Because the prognostic importance of this criterion was unclear, the B rating was not used for staging purposes.<sup>5</sup> Later studies at the NCI and elsewhere indicated that more than 20% atypical (Sézary) lymphocytes was associated with an adverse prognosis in MF,<sup>24,98</sup> although no formal revision to the staging classification was done because this more rigorous B<sub>1</sub> rating was not found to be an independent prognostic variable.<sup>13,24</sup> However, Kim et al have subsequently shown that “significant” blood involvement (> 1000 Sézary cells/mm<sup>3</sup> and/or > 20% Sézary cells) has prognostic significance regardless of T or N rating,<sup>25</sup> and others have corroborated that blood involvement has independent prognostic significance.<sup>61,99,100</sup>

The assessment of blood tumor burden in CTCL based on morphologic features of the neoplastic cells alone (eg, Sézary cell counts) is subjective and prone to interobserver variability, although absolute counts of Sézary cells continue to be used in staging at centers where such counts are routinely performed.<sup>100,101</sup> Blood flow cytometry offers an alternate objective means of identifying and quantifying these neoplastic lymphocytes in the blood. There has been an increasing awareness that neoplastic T cells in CTCL often have altered surface expression of normal markers such as CD3, CD4, CD7, and CD26.<sup>102-108</sup> Deletion of one or more of these markers on the surface of CD4<sup>+</sup> cells is typical of Sézary cells, but the blood of many patients with benign inflammatory dermatoses may also show CD7 deletion.<sup>107,109,110</sup> Loss of CD26 may be a more specific phenotype for the neoplastic lymphocytes.<sup>104,108</sup> Identification of neoplastic cells by flow cytometry is complicated, however, by the fact that all neoplastic lymphocytes may not have the same phenotypic features and several clones may be present in a given patient with SS.<sup>111</sup> The correlation of the percentage of abnormal cells determined by flow cytometry and that by Sézary cell preparation is inexact and may offer differing results in some cases.

Clonal expansion of TCR gene rearrangement in the blood is extremely common in early-stage disease even without a significant population of morphologically or immunophenotypically abnormal cells.<sup>99,112</sup> It is not synonymous with blood involvement by MF/SS since benign lymphoproliferative disorders<sup>113,114</sup> and

**Table 6. Recommended evaluation/initial staging of the patient with mycosis fungoides/Sézary syndrome****Complete physical examination including**

Determination of type(s) of skin lesions

If only patch/plaque disease or erythroderma, then estimate percentage of body surface area involved and note any ulceration of lesions

If tumors are present, determine total number of lesions, aggregate volume, largest size lesion, and regions of the body involved

Identification of any palpable lymph node, especially those  $\geq 1.5$  cm in largest diameter or firm, irregular, clustered, or fixed

Identification of any organomegaly

**Skin biopsy**

Most indurated area if only one biopsy

Immunophenotyping to include at least the following markers: CD2, CD3, CD4, CD5, CD7, CD8, and a B-cell marker such as CD20. CD30 may also be indicated in cases where lymphomatoid papulosis, anaplastic lymphoma, or large-cell transformation is considered.

Evaluation for clonality of TCR gene rearrangement

**Blood tests**

CBC with manual differential, liver function tests, LDH, comprehensive chemistries

TCR gene rearrangement and relatedness to any clone in skin

Analysis for abnormal lymphocytes by either Sézary cell count with determination absolute number of Sézary cells and/or flow cytometry (including CD4<sup>+</sup>/CD7<sup>-</sup> or CD4<sup>+</sup>/CD26<sup>-</sup>)**Radiologic tests**In patients with T<sub>1</sub>N<sub>0</sub>B<sub>0</sub> stage disease who are otherwise healthy and without complaints directed to a specific organ system, and in selected patients with T<sub>2</sub>N<sub>0</sub>B<sub>0</sub> disease with limited skin involvement, radiologic studies may be limited to a chest X-ray or ultrasound of the peripheral nodal groups to corroborate absence of adenopathyIn all patients with other than presumed stage IA disease, or selected patients with limited T<sub>2</sub> disease and the absence of adenopathy or blood involvement, CT scans of chest, abdomen, and pelvis alone  $\pm$  FDG-PET scan are recommended to further evaluate any potential lymphadenopathy, visceral involvement, or abnormal laboratory tests. In patients unable to safely undergo CT scans, MRI may be substituted.**Lymph node biopsy**Excisional biopsy is indicated in those patients with a node that is either  $\geq 1.5$  cm in diameter and/or is firm, irregular, clustered, or fixed

Site of biopsy

Preference is given to the largest lymph node draining an involved area of the skin or if FDG-PET scan data are available, the node with highest standardized uptake value (SUV).

If there is no additional imaging information and multiple nodes are enlarged and otherwise equal in size or consistency, the order of preference is cervical, axillary, and inguinal areas.

Analysis: pathologic assessment by light microscopy, flow cytometry, and TCR gene rearrangement.

some healthy elderly volunteers<sup>115,116</sup> may have clonal TCR gene rearrangements of the blood T cells. Using spectratyping, MF patients even at early stages demonstrate loss of their T-cell repertoire with emergence of one of more clones.<sup>117</sup> The presence of a peripheral blood clone in MF patients, if the same as that in the skin, has been found to have prognostic significance independent of skin stage.<sup>99</sup>

Previously, the ISCL has categorized blood involvement into prognostically significant B ratings based on the degree of involvement, that is, B<sub>0</sub> = absence of significant blood involvement; B<sub>1</sub> = aleukemic, low blood tumor burden; and B<sub>2</sub> = leukemic, high blood tumor burden.<sup>11</sup> The ISCL/EORTC has simplified and clarified the definitions of B<sub>0</sub> to B<sub>2</sub>. B<sub>0</sub> remains 5% or less Sézary cells. B<sub>2</sub> is now defined as a clonal rearrangement of the TCR in the blood and either 1.0 K/ $\mu$ L or more Sézary cells or one of the 2 criteria outlined by the ISCL,<sup>11</sup> that is, (1) increased CD4<sup>+</sup> or CD3<sup>+</sup> cells with CD4/CD8 of 10 or more or (2) increase in CD4<sup>+</sup> cells with an abnormal phenotype ( $\geq 40\%$  CD4<sup>+</sup>/CD7<sup>-</sup> or  $\geq 30\%$  CD4<sup>+</sup>/CD26<sup>-</sup> has been suggested<sup>118</sup>). B<sub>1</sub> is defined as more than 5% Sézary cells but either less than 1.0 K/ $\mu$ L absolute Sézary cells or absence of a clonal rearrangement of the TCR or both.

## Evaluation and staging of the patient with MF/SS

The staging of MF/SS according to the TNMB system implies that an appropriate evaluation of the 4 TNMB systems has been performed. The recommended workup is detailed in Table 6.

The updated ISCL staging classification (Table 7) takes into account the B stage and also differentiates levels of blood involvement (B<sub>1</sub> and B<sub>2</sub>). B<sub>1</sub> is used to separate erythrodermic

patients without overt lymph node involvement (T<sub>4</sub>N<sub>0-2</sub>M<sub>0</sub>) into 2 subgroups, IIIA (T<sub>4</sub>N<sub>0-2</sub>M<sub>0</sub>B<sub>0</sub>) and IIIB (T<sub>4</sub>N<sub>0-2</sub>M<sub>0</sub>B<sub>1</sub>), which will allow determination of the prognostic significance of low blood tumor burden in the setting of erythrodermic CTCL. The ISCL blood rating of B<sub>2</sub> is considered comparable with nodal involvement (N<sub>3</sub> nodal rating). Stage IVA is now defined as any skin stage and either blood involvement (B<sub>2</sub> [IVA<sub>1</sub>]) or nodal lymphoma (N<sub>3</sub> [IVA<sub>2</sub>]) that allows for independent tracking of these 2 important prognostic indicators.

What has not been dealt with adequately in this updated staging is the continued classification of tumor-stage MF at a stage below that of erythroderma with the data from several centers now demonstrating that the survival curves for T<sub>3</sub> patients are similar<sup>24,25,29</sup> or even worse than T<sub>4</sub> patients<sup>53,52,61,84,119,120</sup>. However, it remains unclear to what degree other important prognostic factors such as lymph node or blood involvement or large-cell transformation are influencing these observations. Ideally, comparison of the survival curves of

**Table 7. ISCL/EORTC revision to the staging of mycosis fungoides and Sézary syndrome**

	T	N	M	B
IA	1	0	0	0,1
IB	2	0	0	0,1
II	1,2	1,2	0	0,1
IIIB	3	0-2	0	0,1
III	4	0-2	0	0,1
IIIA	4	0-2	0	0
IIIB	4	0-2	0	1
IVA <sub>1</sub>	1-4	0-2	0	2
IVA <sub>2</sub>	1-4	3	0	0-2
IVB	1-4	0-3	1	0-2

patients with T<sub>3</sub> and T<sub>4</sub> skin ratings who have comparable N and now B ratings should be undertaken before concluding that the hierarchy of T<sub>3</sub> and T<sub>4</sub> should be modified. It is also possible that erythrodermic patients with coexisting tumors may have this important prognostic variable lost in the final staging because only the highest skin T rating is used for staging purposes. For these reasons, the ISCL/EORTC have elected to retain the existing staging parameters until additional information is available.

## Use of the term “stage”

In oncology, the stage assigned to a patient with malignancy at the initial diagnosis and workup is the primary prognostic indicator, and although the condition may go into complete or partial remission, relapse, or progress, the “clinical stage” does not change thereafter.<sup>121</sup> In keeping with this tradition, the formal stage of a patient with MF/SS refers to the overall tumor status at initial diagnosis. However, as with other malignancies, changes in the tumor burden in patients with MF/SS often occur during the course of disease, which affects treatment choices and response to treatment. Moreover, it is important to have a means to indicate both the maximum and the current disease status at the time of enrollment into clinical trials. Therefore, the ISCL/EORTC recommends that, in addition to “clinical stage” at diagnosis of MF/SS patients, that TNMB ratings without the corresponding stage be used to indicate the maximum tumor burden and the current tumor burden. These distinctions will provide a means of communicating the initial, maximum, and current level and type of tumor burden for an individual patient.

## Conclusions

The ISCL/EORTC recommended revisions to the MFCG classification and staging system for CTCL are made both to incorporate advances since 1979 related to tumor-cell biology and diagnostic techniques as pertains to MF and SS and to clarify certain variables that currently impede effective interinstitution and interinvestigator communication and/or the development of standardized clinical

trials in MF and SS. The ISCL/EORTC recognizes that although this revision to the staging and classification of MF and SS further narrows and defines the variables involved, it does not provide a finite staging system that inherently incorporates all potential prognostic factors. There are several current factors, primarily related either to histopathology of lesions (folliculotropic MF, large-cell transformation, patch vs plaque disease) or TCR gene rearrangement in tissue and/or blood, that require further data on their prognostic importance before making formal revisions to the staging system of MF and SS related to them. The revisions to the classification outlined here provide a framework on which to gather data and facilitate validation efforts regarding these variables, all of which can be addressed in an optional fashion for any given MF or SS patient without affecting his/her overall staging. As additional clinical, genetic, or molecular information becomes available, it is anticipated that there will be further revisions to the classification and staging guidelines for MF and SS.

## Authorship

Contribution: E.O. wrote the draft of the paper, participated in paper preparation, and was an active participant in the meetings where revisions to the staging and classification were generated; E.V., N.P., R.W., Y.K., R.K., H.Z., M.D., T.E., S.L., G.W., R.D., A.R., P.H., M.P., M.-G.B., and S.W. participated in paper preparation and were active participants in the meetings where revisions to the staging and classification were generated; G.B. and W.S. participated in paper preparation; L.L. and F.T. were active participants in the meetings where revisions to the staging and classification were generated.

Complete lists of the active members of the International Society for Cutaneous Lymphomas (Document S1) and the European Organization of Research and Treatment of Cancer (Document S2) are available on the *Blood* website; see the Supplemental Appendices link at the top of the online article.

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## References

- Lutzner AM, Edelson R, Schein P, Green I, Kirkpatrick C, Ahmed A. Cutaneous T-cell lymphomas: the Sézary syndrome, mycosis fungoides, and related disorders. *Ann Intern Med*. 1975;83:534-552.
- Lamberg SI, Diamond EL, Lorincz AL, et al. Mycosis fungoides cooperative study. *Arch Dermatol*. 1975;111:457-459.
- Harmer, MH. TNM Classification of Malignant Tumours. 3rd ed. Geneva, Switzerland: UICC—International Union Against Cancer; 1978.
- Oliver HB, Carr DT, Rubin P, et al. American Joint Committee on Cancer: AJCC Cancer Staging Manual. 1st ed. Chicago, IL: Whiting Press; 1978.
- Bunn PA, Lamberg SI. Report of the committee on staging and classification of cutaneous T-cell lymphomas. *Cancer Treat Rep*. 1979;63:725-728.
- Willemze R, Jaffe ES, Burg G, et al. WHO-EORTC classification for cutaneous lymphomas. *Blood*. 2005;105:3768-3785.
- Burg G, Jaffe ES, Kempf W, et al. WHO/EORTC classification of cutaneous lymphomas. In: LeBoit P, Burg G, Weedon D, Sarasin A, eds. WHO Books: Tumors of the Skin. Lyon, France: WHO IARC. Vol X:166-168.
- Wood GS, Weiss LM, Warnke RA, Sklar J. The immunopathology of cutaneous lymphomas: immunophenotypic and immunogenotypic characteristics. *Semin Dermatol*. 1986;5:334-345.
- Burg G, Kempf W, Cozzio A, et al. WHO/EORTC classification of cutaneous lymphomas 2005: histological and molecular aspects. *J Cutan Pathol*. 2005;32:647-674.
- Sentis HJ, Willemze R, Scheffer E. Histopathologic studies in Sézary syndrome and erythrodermic mycosis fungoides: a comparison with benign forms of erythroderma. *J Am Acad Dermatol*. 1986;15:1217-1226.
- Vonderheid EC, Bernengo MG, Burg G, et al. Update on erythrodermic cutaneous T-cell lymphoma: report of the International Society for Cutaneous Lymphomas. *J Am Acad Dermatol*. 2002;46:95-106.
- Diwan AH, Prieto VG, Herling M, Duvic M, Jones D. Primary Sézary syndrome commonly shows low-grade cytologic atypia and an absence of epidermotropism. *Am J Clin Pathol*. 2005;123:510-515.
- Sausville EA, Worsham GF, Matthews MJ, et al. Histologic assessment of lymph nodes in mycosis fungoides/Sézary syndrome (cutaneous T-cell lymphoma): clinical correlations and prognostic import of a new classification system. *Human Pathol*. 1985;16:1098-1109.
- Rappaport H, Thomas LB. Mycosis fungoides: the pathology of extracutaneous involvement. *Cancer*. 1974;34:1198-1229.
- Pimpinelli N, Olsen EA, Santucci M, et al. Defining early mycosis fungoides. *J Am Acad Dermatol*. 2005;53:1053-1063.
- Guitart J, Kennedy J, Ronan S, Chmiel JS, Hsieh Y-C, Variakojis D. Histologic criteria for the diagnosis of mycosis fungoides: proposal for a grading system to standardize pathology reporting. *J Cutan Pathol*. 2001;28:174-183.
- Santucci M, Biggeri A, Feller A, Massi D, Burg G. Efficacy of histologic criteria for diagnosing early mycosis fungoides: an EORTC cutaneous lymphoma study group investigation: European Organization for Research and Treatment of Cancer. *Am J Surg Pathol*. 2000;24:40-50.

18. Massone C, Kodama K, Kerl H, Cerroni L. Histopathologic features of early (patch) lesions of mycosis fungoides: a morphologic study on 745 biopsy specimens from 427 patients. *Am J Surg Pathol*. 2005;29:550-560.
19. Vonderheid EC. On the diagnosis of erythrodermic cutaneous T-cell lymphoma. *J Cutan Pathol*. 2006;33(suppl 1):27-42.
20. Russell-Jones R. Diagnosing erythrodermic cutaneous T-cell lymphoma. *Br J Dermatol*. 2005;153:1-5.
21. Vega F, Luthra R, Medeiros J, Dunmire V, et al. Clonal heterogeneity in mycosis fungoides and its relationship to clinical course. *Blood*. 2002;100:3369-3373.
22. Karenko L, Sarna S, Kähkönen M, Ranki A. Chromosomal abnormalities in relation to clinical disease in patients with cutaneous T-cell lymphoma: a 5-year follow-up study. *Br J Dermatol*. 2003;148:55-64.
23. Zackheim HS, Amin S, Kashani-Sabet M, McMillan A. Prognosis in cutaneous T-cell lymphoma by skin stage: long-term survival in 489 patients. *J Am Acad Dermatol*. 1999;40:418-425.
24. Sausville EA, Eddy JL, Makuch RW, et al. Histopathologic staging at initial diagnosis of mycosis fungoides and the Sézary syndrome: definition of three distinctive prognostic groups. *Ann Int Med*. 1988;109:372-382.
25. Kim YH, Liu HL, Mraz-Gernhard S, Varghese A, Hoppe RT. Long term outcome of 525 patients with mycosis fungoides and Sézary Syndrome: clinical prognostic factors and risk for disease progression. *Arch Dermatol*. 2003;139:857-866.
26. Lamberg SI, Green SB, Byar DP, et al. Clinical staging for cutaneous T-cell lymphoma. *Ann Int Med*. 1984;100:187-192.
27. Zackheim HS, Kashani-Sabet M, Amin S. Topical corticosteroids for mycosis fungoides: experience in 79 patients. *Arch Dermatol*. 1998;134:949-954.
28. LeBoit PE, McCalmont TH. Cutaneous lymphomas and leukemias. In: Elder D, editor. *Lever's Histopathology of the Skin*. Philadelphia, PA: Lippincott-Raven, 1977:805-846.
29. Kashani-Sabet M, McMillan A, Zackheim HS. A modified staging classification for cutaneous T-cell lymphoma. *J Am Acad Dermatol*. 2001;45:700-706.
30. Martí RM, Estrach T, Reverter JC, Mascaró JM. Prognostic clinicopathologic factors in cutaneous T-cell lymphoma. *Arch Dermatol*. 1991;127:1511-1516.
31. Rossiter ND, Chapman P, Haywood IA. How big is a hand? *Burns*. 1996;22:230-231.
32. Amirshaybani HR, Crecelius GM, Timothy NH, Pfeiffer M, Sagers GC, Manders EK. The natural history of the growth of the hand: I, hand area as a percentage of body surface area. *Plast Reconstr Surg*. 2001;107:726-733.
33. Perry RJ, Moore CA, Morgan BDG, Plummer DL. Determining the approximate area of a burn: an inconsistency investigated and re-evaluated [letter]. *Br Med J*. 1996;312:1338.
34. Sheridan RL, Petras L, Basha G, et al. Planimetry study of the percent of body surface represented by the hand and palm: sizing irregular burns is more accurately done with the palm. *J Burn Care Rehabil*. 1995;16:605-606.
35. Lund CC, Browder NC. The estimation of areas of burns. *Surg Gynecol Obstet*. 1944;79:352-358.
36. Epstein EH, Levin DL, Croft JD, Lutzner MA. Mycosis fungoides: survival, prognostic features, response to therapy, and autopsy findings. *Medicine*. 1972;51:61-72.
37. Lamberg SI, Green SB, Byar DP, et al. Status report of 376 mycosis fungoides patients at four years: Mycosis Fungoides Cooperative Group. *Cancer Treat Rep*. 1979;63:701-707.
38. Green SB, Byar DP, Lamberg SI. Prognostic variables in mycosis fungoides. *Cancer*. 1981;47:2671-2677.
39. Edelson RL, Berger C, Gasparro F, et al. Treatment of cutaneous T-cell lymphoma by extracorporeal photochemotherapy: preliminary results. *N Engl J Med*. 1987;316:297-303.
40. Olsen E, Duvic M, Frankel A, et al. Pivotal phase III trial of two dose levels of denileukin diftitox for the treatment of cutaneous T-cell lymphoma. *J Clin Oncol*. 2001;19:376-388.
41. Klemke C-D, Dippel E, Assaf C, et al. Follicular mycosis fungoides. *Br J Dermatol*. 1999;141:137-140.
42. Bonta MD, Tannous ZS, Demierre M-F, Gonzalez E, Harris NL, Duncan LM. Rapidly progressing mycosis fungoides presenting as follicular mucinosis. *J Am Acad Dermatol*. 2000;43:635-640.
43. Gilliam AC, Lessin SR, Wilson DM, Salhany KE. Folliculotropic mycosis fungoides with large cell transformation presenting as dissecting cellulitis of the scalp. *J Cutan Pathol*. 1997;24:169-175.
44. Sangüeza OP. Mycosis fungoides: new insights into an old problem. *Arch Dermatol*. 2002;138:244-246.
45. van Doorn R, Scheffer E, Willemze R. Follicular mycosis fungoides, a distinct disease entity with or without associated follicular mucinosis. *Arch Dermatol*. 2002;138:191-198.
46. Diamandidou E, Colome-Grimmer M, Fayad L, Duvic M, Kurzrock R. Transformation of mycosis fungoides/Sézary Syndrome: clinical characteristics and prognosis. *Blood*. 1998;92:1150-1159.
47. Salhany KE, Cousar JB, Greer JP, Casey TT, Fields JP, Collins RD. Transformation of cutaneous T cell lymphoma to large cell lymphoma: a clinicopathologic and immunologic study. *Am J Pathol*. 1988;132:265-277.
48. Vergier B, de Muret A, Beylot-Barry M, et al. Transformation of mycosis fungoides: clinicopathological and prognostic features of 45 cases. *Blood*. 2000;95:2212-2218.
49. Cerroni L, Rieger E, Hödl S, Kerl H. Clinicopathologic and immunologic features associated with transformation of mycosis fungoides to large cell lymphoma. *Am J Surg Pathol*. 1992;16:543-552.
50. Wood GS, Bahler DW, Hoppe RT, Warnke RA, Sklar JL, Levy R. Transformation of mycosis fungoides: T-cell receptor  $\alpha$  gene analysis demonstrates a common clonal origin for plaque-type mycosis fungoides and CD30+ large-cell lymphoma. *J Invest Dermatol*. 1993;101:296-300.
51. Wolfe JT, Chooback L, Finn DT, Jaworsky C, Rook AH, Lessin SR. Large-cell transformation following detection of minimal residual disease in cutaneous T-cell lymphoma: molecular and in situ analysis of a single neoplastic T-cell clone expressing the identical T-cell receptor. *J Clin Oncol*. 1995;13:1751-1757.
52. Hoppe RT, Cox RS, Fuks Z, Price NM, Bagshaw MA, Farber EM. Electron beam therapy for mycosis fungoides: the Stanford University experience. *Cancer Treat Rep*. 1979;63:691-700.
53. Fuks ZY, Bagshaw MA, Farber EM. Prognostic signs and the management of the mycosis fungoides. *Cancer*. 1973;32:1385-1395.
54. Cheson BD, Horning SJ, Coiffier B, et al. Report of an international workshop to standardize response criteria for non-Hodgkin's lymphomas. *J Clin Oncol*. 1999;17:1244-1253.
55. Kim Y, Willemze R, Pimpinelli N, et al. TNM classification system for primary cutaneous lymphomas other than mycosis fungoides and Sézary syndrome: a proposal of the International Society for Cutaneous Lymphomas (ISCL) and the Cutaneous Lymphoma Task Force of the European Organization of Research and Treatment of Cancer (EORTC). *Blood*. 2007;110:479-484.
56. Burke JS, Colby TV. Dermatopathic lymphadenopathy: comparison of cases associated and unassociated with mycosis fungoides. *Am J Surg Pathol*. 1981;5:343-352.
57. Clendenning WE, Rappaport HW. Report of the committee on pathology of cutaneous T cell lymphomas. *Cancer Treat Rep*. 1979;63:719-724.
58. Scheffer E, Meijer CJLM, van Vloten WA. Dermatopathic lymphadenopathy and lymph node involvement in mycosis fungoides. *Cancer*. 1980;45:137-148.
59. Colby TV, Burke JS, Hoppe RT. Lymph node biopsy in mycosis fungoides. *Cancer*. 1981;47:351-359.
60. Vonderheid EC, Diamond LW, van Vloten WA, et al. Lymph node classification systems in cutaneous T-cell lymphoma: evidence for the utility of the working formulation of non-Hodgkin's lymphomas for clinical usage. *Cancer*. 1994;73:207-218.
61. Toro JR, Stoll HL, Stomper PC, Oseroff AR. Prognostic factors and evaluation of mycosis fungoides and Sézary syndrome. *J Am Acad Dermatol*. 1997;37:58-67.
62. Kern DE, Kidd PG, Moe R, Hanke D, Olerud JE. Analysis of T-cell receptor gene rearrangement in lymph nodes of patients with mycosis fungoides. Prognostic implications. *Arch Dermatol*. 1998;134:158-164.
63. Wechsler J, Diebold J, Gerard-Marchant R. Aspects histopathologiques ganglionnaires au cours des lymphomas cutanés de type mycosis fungoides ou syndrome de Sézary: étude rétrospective de 98 biopsies. *Ann Pathol*. 1997;7:122-129.
64. Lynch JW Jr, Linoila I, Sausville EA, et al. Prognostic implications of evaluation for lymph node involvement by T-cell antigen receptor gene rearrangement in mycosis fungoides. *Blood*. 1992;79:3293-3299.
65. Bakels V, van Oostveen JW, Gordijn RLJ, Walboomers JMM, Meijer CJLM, Willemze R. Diagnostic value of T-cell receptor beta gene rearrangement analysis on peripheral blood lymphocytes of patients with erythroderma. *J Invest Dermatol*. 1991;97:782-786.
66. Wood GS, Haeflner A, Dummer R, Crooks CF. Molecular biology techniques for the diagnosis of cutaneous T-cell lymphoma. *Dermatol Clin*. 1994;12:231-241.
67. Fraser-Andrews EA, Mitchell T, Ferreira S, et al. Molecular staging of lymph nodes from 60 patients with mycosis fungoides and Sézary syndrome: correlation with histopathology and outcome suggests prognostic relevance in mycosis fungoides. *Br J Dermatol*. 2006;155:756-762.
68. Assaf C, Hummel M, Steinhoff M, et al. Early TCR- $\beta$  and TCR- $\gamma$  PCR detection of T-cell clonality indicates minimal tumor disease in lymph nodes of cutaneous T-cell lymphoma: diagnostic and prognostic implications. *Blood*. 2005;105:503-510.
69. Juarez T, Isenbath SN, Polissar NL, et al. Analysis of T-cell receptor gene rearrangement for predicting clinical outcome in patients with cutaneous T-cell lymphoma: a comparison of Southern blot and polymerase chain reaction methods. *Arch Dermatol*. 2005;141:1107-1113.
70. Galindo LM, Garcia FU, Hanau CA, et al. Fine needle aspiration biopsy in the evaluation of lymphadenopathy associated with cutaneous T-cell lymphoma (Mycosis fungoides/Sézary syndrome). *Am J Clin Pathol*. 2000;113:865-871.
71. Papa VI, Hussain HK, Reznick RH, et al. Role of image-guided core-needle biopsy in the management of patients with lymphoma. *J Clin Oncol*. 1996;14:2427-2430.
72. Demharter J, Müller P, Wagner T, Schlimok G, Haude K, Bohndorf K. Percutaneous core-needle biopsy of enlarged lymph nodes in the diagnosis and subclassification of malignant lymphomas. *Eur Radiol*. 2001;11:276-283.
73. Schoengen A, Binder T, Fembacher P, Zeelen U. The fine-needle aspiration cytology of lymph nodes suspected of malignancy: its diagnostic value and capacity to predict the histogenesis. *Dtsch Med Wochenschr*. 1995;120:549-554.

74. Vonderheid EC, Diamond LW, Lai S-M, Au F, Del-lavecchia MA. Lymph node histopathologic findings in cutaneous T-cell lymphoma: a prognostic classification system based on morphologic assessment. *Am J Clin Pathol.* 1992;97:121-129.
75. Breneman DL, Raju US, Breneman JC, et al. Lymph node grading for staging of mycosis fungoides may benefit from examination of multiple excised lymph nodes. *J Am Acad Dermatol.* 2003;48:702-706.
76. Fraser-Andrews E, Whittaker S, Russell-Jones R. Should multiple lymph node biopsies be taken [letter]? *J Am Acad Dermatol.* 2004;51:483.
77. Bunn PA, Huberman MS, Whang-Peng J, et al. Prospective staging evaluation of patients with cutaneous T-cell lymphomas: demonstration of a high frequency of extracutaneous dissemination. *Ann Int Med.* 1980;93:223-230.
78. Lister TA, Crowther D, Sutcliffe SB, et al. Report of a committee convened to discuss the evaluation and staging of patients with Hodgkin's disease: Cotswolds meeting. *J Clin Oncol.* 1989;7:1630-1636.
79. Huberman MS, Bunn PA, Matthews MJ, et al. Hepatic involvement in the cutaneous T-cell lymphomas: results of percutaneous biopsy and peritoneoscopy. *Cancer.* 1980;45:1683-1688.
80. Sibaud V, Beylot-Barry M, Thiébaud R, et al. Bone marrow histopathologic and molecular staging in epidermotropic T-cell lymphomas. *Am J Clin Pathol.* 2003;119:414-423.
81. Graham SJ, Sharpe RW, Steinberg SM, Cotelingam JD, Sausville EA, Foss FM. Prognostic implications of a bone marrow histopathologic classification system in mycosis fungoides and the Sézary syndrome. *Cancer.* 1993;72:726-734.
82. Foss FM, Sausville EA. Prognosis and staging of cutaneous T-cell lymphoma. *Hem Oncolog Clin NA.* 1995;9:1011-1019.
83. Salhany KE, Greer JP, Cousar JB, Collins RD. Marrow involvement in cutaneous T-cell lymphoma: a clinicopathologic study of 60 cases. *Am J Clin Pathol.* 1989;92:747-754.
84. Diamandidou E, Colome M, Fayad L, Duvic M, Kurzrock R. Prognostic factor analysis in mycosis fungoides/Sézary syndrome. *J Am Acad Dermatol.* 1999;40:914-924.
85. Beylot-Barry M, Parrens M, Delaunay M, et al. Is bone marrow biopsy necessary in patients with mycosis fungoides and Sézary syndrome? a histological and molecular study at diagnosis and during follow-up. *Br J Dermatol.* 2005;152:1378-1379.
86. Olsen EA, Delzell E, Jegasothy BV. Second malignancies in cutaneous T-cell lymphoma. *J Am Acad Dermatol.* 1984;10:197-204.
87. Väkevälä L, Pukkala E, Ranki A. Increased risk of secondary cancers in patients with primary cutaneous T cell lymphoma. *J Invest Dermatol.* 2000;115:62-65.
88. Huang KP, Weinstock MA, Clarke CA, Hoppe R, Kim Y. Increased risk for second lymphomas in patients with mycosis fungoides and Sézary syndrome [abstract]. *J Invest Dermatol.* 2005;124:A48.
89. Clendenning WE, Brecher G, Van Scott EJ. Mycosis fungoides: relationship to malignant cutaneous reticulosis and the Sézary syndrome. *Arch Dermatol.* 1964;89:785-792.
90. Lutzner MA, Jordan HW. The ultrastructure of an abnormal cell in Sézary's syndrome. *Blood.* 1968;31:719-726.
91. Duncan SC, Winkelmann RK. Circulating Sézary cells in hospitalized dermatology patients. *Br J Dermatol.* 1978;99:171-178.
92. Meijer CJLM, van Leeuwen AWF, van der Loo EM, van de Putte LBA, van Vloten WA. Cerebriform (Sézary like) mononuclear cells in healthy individuals: a morphologically distinct population of T cells: relationship with mycosis fungoides and Sézary's Syndrome. *Virchows Arch. B Cell Pathol.* 1977;25:95-104.
93. Rosas-Urbe A, Variakojis D, Molnar Z, Rappaport H. Mycosis fungoides: an ultrastructural study. *Cancer.* 1974;34:634-645.
94. Lutzner MA, Hobbs JW, Horvath P. Ultrastructure of abnormal cells in Sézary syndrome, mycosis fungoides and parapsoriasis en plaque. *Arch Dermatol.* 1971;103:375-386.
95. van der Loo EM, Cnossen J, Meijer CJLM. Morphological aspects of T-cell subpopulations in human blood: characterization of the cerebriform mononuclear cells in healthy individuals. *Clin Exp Immunol.* 1981;43:506-516.
96. Yeckley JA, Weston WL, Thorne EG, Krueger GC. Production of Sézary-like cells from normal human lymphocytes. *Arch Dermatol.* 1975;111:29-32.
97. Reinhold U, Herpertz M, Kukul S, Oltermann I, Uerlich M, Kreysel H-W. Induction of nuclear contour irregularity during T-cell activation via the T-cell receptor/CD3 complex and CD2 antigens in the presence of phorbol esters. *Blood.* 1994;83:703-706.
98. Schechter GP, Sausville EA, Fischmann AB, et al. Evaluation of circulating malignant cells provides prognostic information in cutaneous T cell lymphoma. *Blood.* 1987;69:841-849.
99. Fraser-Andrews E, Woolford AJ, Russell-Jones R, Seed PT, Whittaker SJ. Detection of a peripheral blood T cell clone is an independent prognostic marker in Sézary syndrome. *J Inv Dermatol.* 2000;114:117-121.
100. Scarisbrick JJ, Whittaker S, Evans AV, et al. Prognostic significance of tumor burden in the blood of patients with erythrodermic primary cutaneous T-cell lymphoma. *Blood.* 2001;97:624-630.
101. Vonderheid EC, Pena J, Nowell P. Sézary cell counts in erythrodermic cutaneous T cell lymphoma: implications for prognosis and staging. *Leuk Lymphoma.* 2006;47:1841-1856.
102. Kuchnio M, Sausville EA, Jaffe ES, et al. Flow cytometric detection of neoplastic T cells in patients with mycosis fungoides based on levels of T-cell receptor expression. *Am J Clin Pathol.* 1994;102:856-860.
103. Ginaldi L, Matutes E, Farahat N, De Martinis M, Morilla R, Catovsky D. Differential expression of CD3 and CD7 in T-cell malignancies: a quantitative study by flow cytometry. *Br J Haematol.* 1996;93:921-927.
104. Jones D, Dang NH, Duvic M, Washington LT, Huh YO. Absence of CD26 expression is a useful marker for diagnosis of T-cell lymphoma in peripheral blood. *Am J Clin Pathol.* 2001;115:885-892.
105. Haynes B, Mann DL, Hemler ME, et al. Characterization of a monoclonal antibody which defines an immunoregulatory T cell subset for immunoglobulin synthesis in man. *Proc Natl Acad Sci U S A.* 1980;77:2914-2918.
106. Borowitz MJ, Weidner A, Olsen EA, Picker LJ. Abnormalities of circulating T-cell subpopulations in patients with cutaneous T-cell lymphoma: cutaneous lymphoma-associated antigen expression on T cells correlates with extent of disease. *Leukemia.* 1993;7:859-863.
107. Harmon CB, Witzig TE, Katzmann JA, Pittelkow MR. Detection of circulating T cells with CD4+CD7- immunophenotype in patients with benign and malignant lymphoproliferative dermatoses. *J Am Acad Dermatol.* 1996;35:404-410.
108. Bernengo MG, Novelli M, Quaglino P, et al. The relevance of the CD4+CD26- subset in the identification of circulating Sézary cells. *Br J Dermatol.* 2001;144:125-135.
109. Wood GS, Hong SR, Sasaki DT, et al. Leu-8/CD7 antigen expression of CD3+ T cells: comparative analysis of skin and blood in mycosis fungoides/Sézary syndrome relative to normal blood values. *J Am Acad Dermatol.* 1990;22:602-607.
110. Abel EA, Lindae ML, Hoppe RT, Wood G. Benign and malignant forms of erythroderma: cutaneous immunophenotypic characteristics. *J Am Acad Dermatol.* 1988;9:1089-1095.
111. Jackow CM, Cather JC, Hearne V, Asano AT, Musser JM, Duvic M. Association of erythrodermic cutaneous T-cell lymphoma, superantigen-positive *Staphylococcus aureus* and oligoclonal T-cell receptor V $\alpha$  gene expansion. *Blood.* 1997;89:32-40.
112. Muche J, Lukowsky A, Asadullah K, Gellrich S, Sterry W. Demonstration of frequent occurrence of clonal T cells in the peripheral blood of patients with primary cutaneous T-cell lymphoma. *Blood.* 1997;90:1636-1642.
113. Bakels V, van Ostveen JW, Geerts M-L, Gordijn RLJ, et al. Diagnostic and prognostic significance of clonal T-cell receptor beta gene rearrangements in lymph nodes of patients with mycosis fungoides. *J Pathol.* 1993;170:249-255.
114. Weinberg JM, Jaworsky C, Benoit M, Telegan B, Rook AH, Lessin SR. The clonal nature of circulating Sézary cells. *Blood.* 1995;86:4257-4262.
115. Posnett DN, Sinha R, Kabak S, Russo C. Clonal populations of T cells in normal elderly humans: the T cell equivalent to "benign monoclonal gammopathy." *J Exp Med.* 1994;179:609-618.
116. Muche JM, Sterry W, Gellrich S, Rzyany B, Audring H, Lukowsky A. Peripheral blood T-cell clonality in mycosis fungoides and nonlymphoma controls. *Diagn Mol Pathol.* 2003;12:142-150.
117. Yawalkar N, Ferenczi K, Jones DA, et al. Profound loss of T-cell receptor repertoire complexity in cutaneous T-cell lymphoma. *Blood.* 2003;102:4059-4066.
118. Vonderheid EC, Bernengo MG. The Sézary syndrome: hematologic criteria. *Hematol Oncol Clin N Amer.* 2003;17:1367-1389.
119. de Coninck EC, Kim YH, Varghese A, Hoppe RT. Clinical characteristics and outcome of patients with extracutaneous mycosis fungoides. *J Clin Oncol.* 2001;19:779-784.
120. Vonderheid EC, Van Scott EJ, Wallner PE, Johnson WC. A 10-year experience with topical mechlorethamine for mycosis fungoides: comparison with patients treated by total-skin electron-beam radiation therapy. *Cancer Treat Rep.* 1979;63:681-689.
121. Sobin LH, Wittekind Ch. *UICC International Union Against Cancer: TNM Classification of Malignant Tumours.* 6th ed. New York, NY: Wiley-Liss, 2002.